Review

# The use of packaging techniques to maintain freshness in fresh-cut fruits and vegetables: a review

# Mª Alejandra Rojas-Graü, Gemma Oms-Oliu, Robert Soliva-Fortuny & Olga Martín-Belloso\*

Department of Food Technology, TPV-XaRTA, University of Lleida, Rovira Roure 191, 25198 Lleida, Spain

(Received 9 October 2008; Accepted in revised form 11 December 2008)

**Summary** Browning and other discolourations, softening, surface dehydration, water loss, translucency, off-flavour and off-odour development, as well as microbial spoilage are some of the most frequent causes of quality loss in fresh-cut products. Nowadays, the use of innovative modified atmospheres and edible coatings stands out among other techniques in the struggle for maintaining freshness and safety of fresh-cut fruits and vegetables. A few studies have demonstrated the effectiveness of these techniques when applied to different fresh-cut commodities. However, treatment and storage conditions for fresh-cut fruits are still being largely explored to better keep their fresh-like quality attributes. This review discusses the recent advances in the use of innovative modified atmosphere packaging (MAP) systems to maintain freshness of fresh-cut fruits and vegetables. Furthermore, special attention is devoted to the development of coatings that can be used as a complement or alternative to MAP.

Keywords Edible coatings, fresh-cut fruits and vegetables, modified atmosphere packaging.

#### Introduction

Processing of fresh-cut fruits and vegetables usually involves the induction of wounding stresses in the cut tissues as a result of mechanical injury, leading to an increase in their respiration rate (Watada et al., 1996). Respiration involves the oxidative breakdown of complex substrate molecules, normally present in plant cells such as starch, sugars and organic acids to simpler molecules, in the course of which energy, carbon dioxide and water are given out. Atmospheres low in  $Q_2$  (1–5%) and high in  $CO<sub>2</sub>$  (5–10%) have been used to extend the shelf-life of fresh-cut fruits and vegetables by reducing respiration, product transpiration and ethylene production, as  $O_2$  is involved in the conversion of 1-amino-cycloprane-1-carboxylic acid to ethylene (Yang & Hoffman, 1984). In general, an inverse relationship has been shown between respiration rates of fruits and vegetables and their postharvest shelf-life. Reduced  $O_2$  and high  $CO_2$  levels have also been proved to effectively control enzymatic browning, firmness and decay of fresh-cut fruits and vegetables. Besides, the proliferation of aerobic spoilage microorganisms can be substantially delayed with reduced  $O<sub>2</sub>$  levels. However, under certain conditions, the growth of some anaerobic

\*Correspondent: Fax: +34 973 702596; e-mail: omartin@tecal.udl.cat

psychrotrophic pathogens might be allowed or even stimulated (Soliva-Fortuny & Martín-Belloso, 2003a). An adequate  $O<sub>2</sub>$  concentration inside packages is required to limit aerobic respiration without triggering anaerobic processes, as too low  $O_2$  and/or excessive  $CO<sub>2</sub>$  concentrations may induce the production of undesirable metabolites and the unleashing of several physiological disorders (Soliva-Fortuny  $& Martin-Bel$ loso, 2003a). In the last few years, the use of elevated  $O_2$  atmospheres ( $\geq$ 70 kPa  $O_2$ ) has been proposed as an alternative to low  $O<sub>2</sub>$  atmospheres to inhibit the growth of naturally occurring spoilage microorganisms, prevent undesired anoxic respirative processes and maintain the fresh-like quality of fresh-cut produce (Amanatidou et al., 1999; Jacxsens et al., 2001; Van der Steen et al., 2002).

On the other hand, edible coatings can be applied as either a complement or an alternative to modified atmosphere packaging (MAP) to improve the shelf-life of fresh-cut fruits. Edible films and coatings may help to reduce the deleterious effects concomitant with minimal processing, not solely retarding food deterioration and enhancing its quality, but also improving its safety because of their natural biocide activity or by incorporating antimicrobial compounds (Petersen et al., 1999). In fact, the application of edible coatings to deliver active substances is one of the major advances reached so far to increase the shelf-life of fresh-cut produce.

doi:10.1111/j.1365-2621.2009.01911.x  $\circledcirc$  2009 The Authors. Journal compilation  $\circledcirc$  2009 Institute of Food Science and Technology

This review highlights some of the most recent findings regarding the use of innovative packaging systems to maintain freshness of fresh-cut fruits and vegetables. An overview of studies on MAP and edible coatings used for preserving fresh-cut fruits and vegetables is provided.

## Maintaining the microbiological stability of fresh-cut fruits and vegetables

Packaging under appropriate atmosphere conditions can effectively control the growth of microorganisms on the surface of fresh-cut fruits. The proliferation of aerobic microorganisms can be substantially delayed with reduced  $O<sub>2</sub>$  levels. The growth of Gram-negative aerobes such as Pseudomonas is specially inhibited, more than for Gram-positive, microaerophilic species such as Lactobacillus. High  $CO<sub>2</sub>$  concentrations are also generally effective in controlling the growth of most aerobic microorganisms, specificallyGram-negative bacteria andmoulds, but fail to inhibit most yeasts (Al-Ati & Hotchkiss, 2002). On fresh-cut apples, pears and melon, microbial growth was not inhibited under atmosphere containing moderately high  $CO_2$  concentrations (2.5 kPa  $O_2$  + 7 kPa  $CO_2$ ); indeed, the growth of fruit bacterial populations appeared to be at least as fast as under atmospheric conditions (Soliva-Fortuny & Martín-Belloso, 2003b; Soliva-Fortuny et al., 2004b; Oms-Oliu et al., 2008b, 2008c). Low  $O_2$ concentrations inhibit the growth of the aerobic microbiota but can also stimulate the proliferation of anaerobic psychrotrophic microorganisms. Too low  $O_2$  concentrations inside packages of fresh-cut vegetables may pose a safety risk, as the growth of anaerobic foodborne pathogens might be allowed or even stimulated (Farber, 1991). Potential foodborne pathogens, such as Yersinia enterocolitica, Aeromonas hydrophila, A. caviae and Listeria monocytogenes have been isolated from packaged lettuce (Szabo *et al.*, 2000). Restrictive  $O_2$  atmospheres have also been shown to stimulate the growth of L. monocytogens (Conway et al., 2000; Corbo et al., 2005), E. coli (Gunes & Hotchkiss, 2002) or Clostridium botulinum (Larson & Johnson, 1999). In addition, lactic acid bacteria can grow under moderate  $CO<sub>2</sub>$  environments (Al-Ati & Hotchkiss, 2002). Therefore, an appropriate combination of gas composition, package dimensions and permeability adapted to the respiration of the product is critical to reach a sustainable equilibrium of gas concentrations. This equilibrium must ensure that  $O_2$  levels inside the packages are high enough to avoid the triggering of anaerobic fermentative processes (Martín-Belloso et al., 2007).

Recently, some researchers have claimed that superatmospheric  $O_2$  concentrations ( $\geq$ 70 kPa) can be an alternative to low  $O_2$ -modified atmospheres to prevent undesired anoxic respiration, inhibit the growth of naturally occurring spoilage microorganisms, and maintain fresh-like sensory quality of fresh-cut produce (Amanatidou et al., 1999; Jacxsens et al., 2001; Van der Steen et al., 2002). Results by Allende et al. (2004) are consistent with these claims and show that high  $O_2$ atmospheres (80–100 kPa) alleviated tissue injury, reduced microbial growth and were beneficial in maintaining quality of fresh-cut baby spinach (Allende et al., 2004). However, reports on the effect of high  $O<sub>2</sub>$ concentrations on the growth of the aerobic microbiota on fresh-cut mixed salads were inconclusive (Allende et al., 2002). Lactic acid bacteria and Enterobacteriaceae appeared to be inhibited under high  $O<sub>2</sub>$  concentrations but, on the other hand, the growth of yeasts and Aeromonas caviae was stimulated under high  $O<sub>2</sub>$  levels, whereas psychrotrophic bacteria and L. *monocytogenes* were not affected. Wszelaki & Mitcham (2000) found that 80–100 kPa  $O_2$  inhibited the growth of *Botrytis* cinerea on strawberries. Consistently, an initial atmosphere of  $\geq 70$  kPa O<sub>2</sub> has been reported to retard the growth of moulds (Van der Steen et al., 2002) and yeasts (Jacxsens et al., 2003) on strawberries and raspberries. In fresh-cut pears, Candida parapsilosis survived on inoculated samples stored under 70 kPa  $O_2$ , whereas Rhodotorula mucilaginosa was shown to be sensitive to superatmospheric  $O_2$  concentrations (Oms-Oliu *et al.*, 2008b). Rhodotorula yeast genera found in fresh-cut 'Piel de Sapo' melon and 'Nam Dokmai' mango cubes were also inhibited when exposed to high  $O<sub>2</sub>$  concentrations (Poubol & Izumi, 2005; Oms-Oliu et al., 2008c). The growth of aerobic psychrophilic microorganisms on fresh-cut pear and melon was significantly reduced under 70 kPa  $O_2$  and 2.5 kPa  $O_2$  + 7 kPa  $CO_2$  atmospheres in comparison to passive MAP (Table 1) Oms-Oliu et al. (2008b, 2008c). Consistently, Jacxsens et al. (2001) could not report differences in the growth of psychrotrophic microorganisms on chicory endives stored for 1 week under conventional atmospheres  $(3 \text{ kPa } O_2 \text{ and } 5 \text{ kPa } CO_2)$  and superatmospheric  $O_2$ concentrations (95 kPa  $O_2$  and 5 kPa  $CO_2$ ). Amanatidou *et al.* (1999) evaluated the potential of high  $O_2$ concentrations to inhibit the growth of a number of microorganisms and pathogenic bacteria typically associated with minimally processed vegetables. They found a more pronounced inhibitory effect on bacterial growth when high  $O_2$  concentrations (80 and 90 kPa) were combined with high  $CO<sub>2</sub>$  concentrations (10 and 20 kPa). According to these authors, high  $O_2$  concentrations can generate reactive oxygen species (ROS) that damage vital cell components and thereby reduce microbial growth when oxidative stresses overwhelm cellular antioxidant protection systems. However, the sensitivity of different organisms to  $O_2$  may greatly vary. Some of them could have developed strategies to avoid lethal damage, by inducing enzyme activities that decrease the accumulation of injurious ROS (Kader & Ben-Yehoshua, 2000).

Table 1 Growth of aerobic psychrophilic microorganisms and yeast and moulds on fresh-cut 'Flor de Invierno' pear and 'Piel de Sapo' melon stored under 2.5 kPa O<sub>2</sub> and 7 kPa CO<sub>2</sub>, 70 kPa O<sub>2</sub> and passive modified atmosphere packaging over 14 days at 4 °C (Source: Oms-Oliu et al., 2008b,c)

Day	2.5 kPa $O_2$ + 7 kPa $CO_2$				Air				70 kPa O <sub>2</sub>			
	<b>PS</b>		YM		<b>PS</b>		YM		<b>PS</b>		YM	
	<b>Melon</b>	Pear	<b>Melon</b>	Pear	<b>Melon</b>	Pear	<b>Melon</b>	Pear	Melon	Pear	<b>Melon</b>	Pear
0	1.2a	2.1a	1.7a	3.2a	1.4a	2.3a	1.5a	3.2a	1.5a	2.1a	1.5a	3.2a
3	2.3 <sub>b</sub>	2.8 <sub>b</sub>	1.6a	3.5 <sub>b</sub>	3.5 <sub>b</sub>	3.6 <sub>b</sub>	2.2 <sub>b</sub>	3.6 <sub>b</sub>	2.2 <sub>b</sub>	2.6 <sub>b</sub>	2.0 <sub>b</sub>	3.1a
	4.1c	3.9 <sub>c</sub>	2.5 <sub>b</sub>	3.5 <sub>b</sub>	6.2c	4.6c	3.6 <sub>c</sub>	3.4ab	4.2c	3.8 <sub>c</sub>	2.3 <sub>b</sub>	3.1a
10	5.9d	4.3 <sub>d</sub>	2.8 <sub>b</sub>	3.8 <sub>c</sub>	7.3d	6.1 <sub>d</sub>	4.1 <sub>d</sub>	3.6 <sub>b</sub>	6.8d	3.9 <sub>c</sub>	2.7c	2.9a
14	7.8e	5.7e	2.9 <sub>b</sub>	3.7 <sub>c</sub>	7.9d	7.0e	4.7 <sub>d</sub>	4.0c	7.6e	5.1 <sub>d</sub>	2.7c	2.9a

Values within a column followed by the same small setter indicate that mean values are not significantly different by Duncan's multiple-range test  $(P < 0.05)$ 

Pear wedges were dipped into N-acetylcysteine (0.75% w/v) and glutathione (0.75% w/v) solution. Aerobic psychrophilic microorganisms (PS); yeast and moulds (YM).

The use of edible coatings with antimicrobial properties or with incorporation of antimicrobial compounds is another potential alternative to enhance the safety of fresh-cut produce. Application of antimicrobial agents directly on the food surface may have limited benefits because the active substances are neutralised on contact with the surface or diffuse rapidly from the surface into the product (Min & Krochta, 2005). Antimicrobial edible films and coatings provide more inhibitory effects against spoilage and pathogenic bacteria by maintaining effective concentrations of the active compounds on the food surfaces (Gennadios & Kurth, 1997). In fact, edible films can be designed to slow down the diffusion of antimicrobial substances from the food surface (Dawson et al., 2002; Sebti et al., 2002). Several types of edible coatings have been used for extending shelf-life of fresh commodities. For instance, chitosan, a film-forming polysaccharide, has been widely used because of their ability of inhibiting the growth of many pathogenic bacteria and fungi (Romanazzi et al., 2002). Chien et al. (2007) reported the effectiveness of chitosan in maintaining quality and extending shelf-life of sliced mango. Assis & Pessoa (2004) and Han et al. (2005) also proposed chitosan for extending the shelf-life of sliced apples and fresh strawberries respectively. Park et al.  $(2005)$  reported a reduction of 2.5 and 2 log CFU g<sup>-1</sup> in the counts of Cladosporium sp. and Rhizopus sp., respectively, on strawberries coated with a chitosanbased edible film, just after the coating application. A reduction in the counts of aerobic and coliform microorganisms was also observed during storage. Eissa (2007) reported that the application of a chitosan-based coating on fresh-cut mushrooms contributes to the extension of their shelf-life, in maintaining quality and, to some extent, in controlling physiological decay. Durango et al. (2006) reported that the use of a coating containing chitosan and yam starch is a viable alterna-

tive for controlling microbiological growth on minimally processed carrots. Other edible coatings with antimicrobial properties have been used on fresh-cut produce. Recently, some authors have proposed the use of Aloe vera gels as antimicrobial coatings for fruits and vegetables, because of their proven antifungal activity (Martínez-Romero et al., 2003; Jasso de Rodríguez et al., 2005). Valverde et al.  $(2005)$  and Martinez-Romero et al. (2006) proposed Aloe vera gel-based edible coatings for preventing moisture loss, reducing texture decay and controlling respiratory rate of table grapes and sweet cherries, respectively, while reducing microbial proliferation. Lee et al. (2003) extended the shelf-life of refrigerated apple slices by more than 2 weeks, when using a coating containing carrageenan, ascorbic acid, citric acid and oxalic acid. Amanatidou et al. (2000) observed a reduction of at least 2 log C- $FU$  g<sup>-1</sup> in the natural microbiota of minimally processed carrots when calcium chloride and citric acid treatments where combined with a sodium alginate edible coating prior to packaging under modified atmospheres.

Edible films and coatings carrying antimicrobial compounds provide an innovative way to improve the safety and shelf-life of food systems. The incorporation of antimicrobial agents into edible coatings is gaining importance as potential treatments to reduce the populations of deleterious microorganisms. Antimicrobials that are suitable for incorporation into edible films and coatings can be grouped into several categories, including organic acids (acetic, benzoic, lactic, propionic and sorbic), fatty acid esters (glyceryl monolaurate), polypeptides (lysozyme, peroxidase, lactoferrin and nisin), plant essential oils (cinnamon, oregano and lemongrass), nitrites and sulphites (Franssen & Krochta, 2003). Numerous studies have demonstrated that antimicrobial edible films can reduce bacterial levels on

many products. Rojas-Graü et al. (2007a) reported the efficacy of alginate and gellan edible coatings with the antimicrobial effect of plant essential oils (lemongrass, oregano oil and vanillin) to prolong shelf-life of fresh-cut apples. In line with these studies, Raybaudi-Massilia et al. (2008a) observed that the addition of cinnamon, clove or lemongrass oils or their active compounds into an alginate-based coating increased its antimicrobial effect, reduced the population of E. coli O157:H7 by more than 4 log CFU  $g^{-1}$  and extended the microbiological shelf-life of 'Fuji' apples for at least 30 days. Raybaudi-Massilia et al. (2008b) also evaluated the ability of an alginate-based coating carrying malic acid and essential oils (cinnamon, palmarosa and lemongrass) to improve the shelf-life and safety of fresh-cut melon. According to their results, the incorporation of 0.3% palmarosa oil into the coating looks promising, since it was accepted by panelists, maintained the fruit quality parameters, inhibited the growth of the native microbiota and reduced the population of inoculated Salmonella enteritidis. Chen et al. (1999) developed a methylcellulose edible coating as carrier of benzoic acid to inhibit the growth of osmophilic yeasts on Taiwanese-style fruits (Ten-shing mei and Ching-shuan mei). Yeasts growth was inhibited when both fruits contained  $50-100 \mu$ g benzoic acid per gram. Zhuang et al. (1996) showed that hydroxypropyl methylcellulose coatings containing ethanol were effective in inactivating Salmonella Montevideo on the surface of fresh tomatoes. Latter, Franssen & Krochta (2003) studied the effect of the incorporation of citric, sorbic and acetic acid into HPMC coatings cast on tomatoes and observed that the application of the coating alone resulted in a 2 log  $CFUg^{-1}$  reduction of Salmonella Montevideo, whereas the addition of 0.4% sorbic acid to the coating led to a further significant reduction. Likewise, García et al. (1998) reduced the microbial growth and extended the storage life of fresh strawberries using a starch-based coating containing potassium sorbate.

# Preserving quality loss of fresh-cut fruits and vegetables

Processing operations can induce undesirable changes in colour and appearance during storage and marketing. Browning, degradation and oxidation of pigments, water loss, translucency, whitening or surface dehydration are likely to occur as a consequence of wounding.

#### Browning

Most strategies to control cut-edge browning have focused on theoretical approaches to modulate polyphenol oxidase (PPO) enzyme activities (Martinez & Whitaker, 1995). Low  $O_2$  concentrations (0.25–5%) in combination with moderate levels of  $CO<sub>2</sub>$  (10–20%) have been used to maintain the visual appearance of several fresh-cut fruits such as peach (Palmer-Wright & Kader, 1997; Gorny et al., 1999), kiwifruit (Agar et al., 1999), mango (Rattanapanone et al., 2001) and melon (Qi et al., 1999; Bai et al., 2001; Oms-Oliu et al., 2007a). The destruction of fruit cellular compartmentation allows the oxidation of phenolic compounds by PPO, thus originating colourless quinones that later on polymerise forming melanins (Gil et al., 2006). Because the  $O_2$  is needed for browning reactions, MAP with low  $O_2$  and high  $CO_2$  levels can contribute positively to avoid browning in fresh-cut produce. However, low  $O<sub>2</sub>$ and elevated  $CO<sub>2</sub>$  atmospheres can not effectively inhibit browning of fresh-cut fruits and vegetables such as apple, banana, pear, potato or artichoke, because of their high phenolic content. MAP systems in combination with antioxidant treatments have been suggested to delay browning of fresh-cut apples (Rojas-Graü *et al.*, 2007b, 2008a), pears (Oms-Oliu *et al.*, 2008a), mangoes (González-Aguilar et al., 2000) and bananas (Vilas-Boas & Kader, 2006). Among such compounds, ascorbic acid is very effective in inhibiting enzymatic browning, primarily because of its ability to reduce quinones to phenolic compounds before they undergo further reaction to form pigments (Iyengar & McEvil, 1992). Some post-cutting dips of N-acetyl-L-cysteine and/or reduced glutathione in combination with a 2.5 kPa  $O_2$  + 7 kPa  $CO<sub>2</sub>$  atmosphere have been proved to effectively prevent browning of fresh-cut apples and pears (Rojas-Grau et al., 2007b; Oms-Oliu et al., 2008a). The appearance of browning in some leaf vegetables, e.g. lettuce, can be attributed to the wounding response, which may induce biosynthesis of phenolic compounds, which are substrates of PPO. It has been shown that high  $CO<sub>2</sub>$ atmospheres could inhibit the biosynthesis of phenolic compounds (Ke & Saltveit, 1989). However, an excessive restriction of the  $O_2$  levels surrounding a fruit or vegetable in a modified atmosphere package could lead to internal decay and off-flavour formation. The application of high  $O_2$  atmospheres has been suggested as an alternative to low  $O_2$  and high  $CO_2$  concentrations to inhibit enzymatic browning. Day (1996) hypothesised that high  $O<sub>2</sub>$  concentrations may cause substrate inhibition of PPO or alternatively, high levels of colourless quinones may cause feedback inhibition of the enzyme. Atmospheres containing 80 kPa  $O_2$  and 20 kPa  $CO_2$ have been recommended to inhibit PPO activity and to control the enzymatic browning in 'Iceberg' lettuce (Heimdal et al., 1994). However, the results achieved in this field are often controversial. In some commodities, high  $O<sub>2</sub>$  concentrations alone cannot effectively prevent browning of fresh-cut produce. High  $O_2$  partial pressures (100 kPa) have been shown to have some positive effects on colour only when combined with an adequate dip of ascorbic and citric acids (Limbo & Piergiovanni, 2006). Gorny et al.  $(2002)$  also found that low-O<sub>2</sub>  $(0.25)$ or 0.5 kPa), elevated- $CO<sub>2</sub>$  (4, 10 or 20 kPa  $CO<sub>2</sub>$ ) or high- $O<sub>2</sub>$  (40, 60 or 80 kPa) atmospheres alone did not effectively prevent surface browning of fresh-cut pear slices. Hence, the combined use of a dip of N-acetylcysteine and glutathione at 0.75% prior to packaging under 70 kPa  $O<sub>2</sub>$  effectively maintained the colour of fresh-cut pears during storage (Oms-Oliu et al., 2008a).

Coatings based on edible compounds from several sources have been evaluated to maintain the visual appearance of fresh-cut fruits and vegetables. Brancoli & Barbosa-Cánovas (2000) decreased surface discolouration of apple slices by coating slices with maltodextrin, methylcellulose ascorbic acid and calcium chloride. Le Tien et al. (2001) achieved reduced browning rates in apple and potato slices coated with a combination of whey protein and carboxymethyl cellulose. Perez-Gago et al. (2003b, 2005) inhibited browning of apple slices by using composite coatings prepared from whey protein isolate or concentrate and beeswax or carnauba wax. Sonti *et al.* (2003) coated apple cubes with whey protein concentrate and whey protein isolate, obtaining a delay in browning and texture decay. With this same purpose, McHugh & Senesi (2000) proposed a coating made of a mixture of apple purée, pectin and vegetable oils. The addition of a lipid in the coating formulation remarkably restricted the gas transfer through the forming matrix and improved protection against the outer atmosphere. Shon & Haque (2007) observed a decrease in browning of cut apples and potatoes when using an edible coating containing sour whey flour. They also concluded that the coating effect was not at all beneficial in preventing browning of onion slices. Pen & Jiang (2003) reported that a chitosan edible coating applied on fresh-cut Chinese water chestnuts retarded the development of browning, maintained sensory quality and retained levels of total soluble solids, acidity and ascorbic acid in coated slices.

In addition, antioxidants can be added into the coating matrix to protect the cut surface against oxidative rancidity, degradation and enzymatic browning. Olivas et al. (2003) preserved fresh-cut pear wedges from surface browning by applying a methylcellulosebased coating containing ascorbic and citric acids. Similar results were obtained by Nisperos-Carriedo et al. (1992) who incorporated ascorbic acid into edible coatings to reduce enzymatic browning in whole and sliced mushrooms. Rojas-Graü et al. (2007c) and Tapia et al. (2005) applied alginate- and gellan-based coatings with the addition of cysteine, glutathione and ascorbic acid to fresh-cut apples and papayas, proving that such coatings are good carriers of antioxidant agents. Likewise, Perez-Gago et al. (2006) reduced browning of cut apples by using a whey protein concentrate-beeswax coating containing ascorbic acid, cysteine, or 4-hexylresorcinol. Lee et al. (2003) studied the effect of carrageenan and whey protein concentrate edible coatings in combination with anti-browning agents on fresh-cut apple slices and observed that the incorporation of ascorbic, citric and oxalic acids was advantageous in maintaining colour during 2 weeks.

# Discolouration

Discolouration is a particular problem in vegetable products such as onion, garlic and leek, which can develop pink, red, green, bluegreen or blue tonalities as a consequence of cell disruption (Toivonen & Brummell, 2008). This kind of quality loss is more frequent in fresh produce than in fresh-cut commodities, but can develop in cases where cutting operations or handling lead to significant tissue damage (Howard et al., 1994). For instance, pink discolouration, which is one of the major causes of quality loss in fresh-cut lettuce, was effectively controlled by flushing lettuce pieces with  $\geq 2.5$  kPa O<sub>2</sub> concentrations (Kim *et al.*, 2005). However,  $CO<sub>2</sub>$  accumulation (5–15 kPa) has been shown to cause the development of brown stain (Lipton, 1987), which appears first on the midrib surfaces of some leafy vegetables as a variable-sized sunken or yellowed area that can later become more delineated and brown (López-Gálvez et al., 1996). In fresh-cut carrots or tomatoes, a whitish surface appearance has been attributed to an enzyme-stimulated reaction related to dehydration of surfaces or formation of the wound-induced lignin barrier (Cisneros-Zevallos et al., 1995; Artés *et al.*, 1999). Neither  $1\%$  O<sub>2</sub> +  $10\%$  CO<sub>2</sub> nor 50%  $O_2$  + 30%  $CO_2$  atmospheres avoided the development of white discolouration on fresh-cut carrots after 12 days (Amanatidou et al., 2000). These authors combined the use of an alginate-based coating with MAP to avoid this disorder. In green vegetables, the senescence process usually leads to a yellow colouration of the tissue, because of the degradation of chlorophylls and the formation of pheophytins. The maintenance of refrigeration temperatures and a high relative humidity, combined with atmospheres lowered in  $O_2$  and moderately enriched in  $CO<sub>2</sub>$ , are shown to delay chlorophylls degradation (Artés et al., 2007). In shredded Galega kale, yellow discolouration because of chlorophylls degradation was much higher in air than under low  $O<sub>2</sub>$ atmospheres  $(1-3 \text{ kPa } O_2)$  (Fonseca *et al.*, 2005).

Edible coatings may be effectively used to maintain the bright orange colour of fresh-cut carrots during storage, by reducing the incidence of ''white blush'' as consequence of the surface dehydration of cut surfaces (Bolin & Huxsoll, 1991). White discolouration in carrots can be avoided with the use of an appropriate edible coating. Howard & Dewi (1995) and Li & Barth (1998) highlighted the positive effect of an edible cellulosebased coating on the development of white surface discolouration in minimally processed carrots. Avena-

Bustillos et al. (1994a) and Mei et al. (2002) reported similar results when coating carrots with an emulsion of sodium caseinate/stearic acid emulsion and a xanthan gum solution respectively.

## **Translucency**

Another factor with a great impact on the appearance of fresh-cut products is the change in homogeneity of the tissue that results in the development of translucency after processing. This physiological disorder, characterised by dark and glassy flesh, seems to be of particular importance in tomatoes (Artés et al., 1999; Gil et al., 2002; Lana et al., 2006), melons (Aguayo et al., 2004a; Saftner et al., 2005), papayas (O'Connor-Shaw et al., 1994) pears (Soliva-Fortuny et al., 2002b) and pineapples (Chen & Paull, 2001; Montero-Calderón et al., 2008). Development of translucency has been found to be the principal visual sign of deterioration in MAPpackaged fresh-cut 'Piel de Sapo' melon (Aguayo et al., 2003, 2004a; Bai et al., 2001, 2003). It has been reported that 15 kPa  $CO<sub>2</sub>$  reduced decay and translucency in 'Honeydew' and 'Cantaloupe' melon pieces (Portella & Cantwell, 1998). A 2.5 kPa  $O_2$  + 7 kPa  $CO_2$  atmosphere best maintained the visual quality of fresh-cut 'Piel de Sapo' melon compared with passive MAP and 70 kPa  $O_2$  atmospheres for 2 weeks at 4 °C. Translucency of watersoaked areas is also the most frequent disorder in fresh-cut tomatoes stored under MAP (Gil et al., 2002), even observed at low temperatures, and has been considered a chilling injury symptom (Hong & Gross, 2000). Fresh-cut pepper and cucumber were sensitive to chilling injury during storage at  $5^{\circ}$ C (Kang & Lee, 1997). Fresh-cut tomato also exhibited slight symptoms of chilling injury when stored at  $0^{\circ}C$ (Aguayo et al., 2004b).

As far as we know, there is little information about the application of edible coatings to prevent translucency in fresh-cut produce. Recently, Oms-Oliu et al. (2008e) reported a substantial decrease in the whiteness index of fresh-cut melon coated with a gellan-based coating, which could be linked to an increase in translucency. In the same way, Sothornvit & Rodsamran (2006) observed an important increase in translucency of fresh-cut mango coated with a mango edible film, which was higher when cut fruits were stored at room temperature.

## Softening

Many of the changes occurring in fresh-cut products are influenced by the effect of processing operations, which often result in a dramatic firmness loss.  $O_2$  and  $CO_2$ concentrations do not usually have much effect on the texture of fresh-cut fruits. Atmospheres of  $2$  kPa  $O_2$ , 4 kPa  $O_2$ , 4 kPa  $CO_2$  and 10 kPa  $CO_2$  did not affect the softening rate of fresh-cut banana slices stored for 3 days at 10 °C. However, when low  $O_2$  levels (2 and 4 kPa  $O_2$ ) were combined with high  $CO_2$  concentrations (5 and 10 kPa) softening was slightly enhanced (Vilas-Boas & Kader, 2006). Similarly, Gorny et al. (2002) reported that low  $O_2$  (0.25 or 0.5 kPa) and elevated  $CO_2$ (5, 10 or 20 kPa) atmospheres alone did not effectively prevent softening of fresh-cut pear slices. On the other hand, kiwifruit slices exhibited delayed softening when stored in an ethylene-free atmosphere of  $2-4$  kPa  $O<sub>2</sub>$ and/or 5–10 kPa  $CO<sub>2</sub>$  (Agar *et al.*, 1999). However, the use of dips, containing calcium salts, is usually more effective than MAP in preserving fresh-cut fruits from softening (Oms-Oliu et al., 2007a). Oms-Oliu et al. (2008b) related firmness loss in fresh-cut pears to excessively low  $O_2$  and high  $CO_2$  concentrations. Soliva-Fortuny et al. (2002b, 2002c) showed that high  $CO<sub>2</sub>$ concentrations induce tissue breakdown and formation of important amounts of exudates, evidenced as droplets on the cellular surface and flooded intercellular spaces of fresh-cut pear and apple tissues (Soliva-Fortuny et al. (2002b, 2002c). The use of high  $O<sub>2</sub>$  levels to prevent anaerobic processes was shown to have a positive effect on maintaining firmness of fresh-cut 'Piel de Sapo' melon (Oms-Oliu et al., 2008c).

Texture loss can also be prevented with the use of an appropriate edible coating. Del-Valle et al. (2005) improved the texture of strawberries using a cactusmucilage edible coating. Maftoonazad & Ramaswamy (2004) observed improved firmness in avocados coated with methylcellulose-based edible coating during storage. Texture enhancers such as calcium chloride may be added to edible coatings to improve quality during storage of fresh-cut produce. Hence, the use of calcium chloride for crosslinking some polymers, could minimise softening phenomena. Rojas-Graü et al. (2007c) observed that apple wedges coated with alginate and gellan edible coatings maintained their initial firmness during refrigerated storage, corroborating that both coatings are good carriers of firming agents such as calcium chloride, which is used for crosslinking the polymers. Similar results were obtained by Lee et al. (2003) who indicated that incorporating 1% of calcium chloride within the whey protein concentrate coating formulation helped to maintain firmness of fresh-cut apples pieces. Olivas et al. (2007) maintained firmness of fresh-cut 'Gala' apples during storage by using an alginate-based coating after immersion into a calcium dip. Likewise, Han *et al.* (2004) reported that a chitosan-based coating containing calcium resulted in at least a 24% reduction in the drip loss of frozen-thawed raspberries and increased their firmness by about 25% in comparison with uncoated fruits.

The other major factors affecting texture are water loss and osmotic changes (Saladié et al., 2007). Water loss can be greatly retarded by appropriate packaging



Figure 1 Effect of packaging conditions on juice leakage volume of fresh-cut pineapple stored at 5 °C. High O<sub>2</sub>: 38–40%; low O<sub>2</sub>: 10–12%; edible coating: alginate (2%). Data shown are mean  $\pm$  standard deviation. Duncan's test at  $P \le 0.05$  (*Source*: Montero-Calderón et al., 2008).

(Toivonen & Brummell, 2008). For instance, edible coatings have been extensively used to protect fresh-cut fruit from surface dehydration. The coating acts as a gas barrier around each fruit piece and creates a modified atmosphere inside each coated piece (Rojas-Grau¨ & Martín-Belloso, 2008b). Avena-Bustillos et al. (1994b, 1997) reduced water loss of apples, celery sticks and zucchini using an emulsion containing calcium caseinate and an acetylated monoglyceride. Montero-Calderón et al. (2008) reported that the use of alginate coatings significantly improved shelf-life of fresh-cut pineapple, as reflected in higher juice retention in contrast with the substantial juice leakage observed in other evaluated packaging conditions (Fig. 1). McHugh & Senesi (2000) significantly reduced moisture loss of fresh-cut apples when applying wraps made from apple puree containing various concentrations of lipids. Similarly, Wong et al. (1994) reduced between 12 and 14 times water loss of apple slices by coating them with a cellulose/lipid bilayer edible film.

# Keeping the nutritional composition and antioxidant potential of fresh-cut fruits and vegetables

Currently, there are a few works reporting the effect of MAP on nutritional composition and bioactive compounds of fresh-cut fruit and vegetables. Some of these works have reported the effect of MAP on the sugar and organic acids content of fresh-cut fruits and vegetables. Namely, Carlin et al. (1990) found good retention of d-sucrose in fresh-cut carrots stored under 10% to 40%  $CO<sub>2</sub>$  with  $2\%$  or  $10\%$   $O<sub>2</sub>$ . Depletion of D-sucrose concentrations was also significantly retarded when an alginate-based coating was combined with 20% or 30% CO<sub>2</sub> (Amanatidou et al., 2000). However, González-Aguilar et al. (2000) did not find any significant effect of packaging conditions on the sugars and organic acids content of fresh-cut mangoes. On the other hand, an appropriate combination of MAP with refrigeration temperatures may help to maintain vitamin C concentrations by limiting ascorbic acid oxidation. Lamikanra et al. (2000) concluded that storage temperature is the most influential factor affecting the organic acids content in fresh-cut melon. Ascorbic acid of fresh-cut bell peppers stored under MAP for 15 days was better maintained at  $5^{\circ}$ C than at 10 °C (González-Aguilar et al., 2004). Furthermore, the  $O<sub>2</sub>$  availability in the package headspace has a dramatic effect on the vitamin C decrease throughout storage. According to Soliva-Fortuny & Martín-Belloso (2003b), the vitamin C content of fresh-cut pears was kept almost constant throughout storage in the absence of  $O_2$ . Barth *et al.* (1993) reported significantly higher amounts of ascorbic acid in broccoli stored in MAP than in unpacked broccoli. Ascorbic acid degradation of shredded Galega kale was also more pronounced in air than under hypoxic environments, although almost no differences were found between 1, 2 and 3 kPa  $O_2$  atmospheres (Fonseca et al., 2005). Consistently, the vitamin C content of fresh-cut spinach was found to be better maintained for 7 days under  $6\%$  O<sub>2</sub> + 14% CO<sub>2</sub> than in air. However, Gil et al. (1999) found that ascorbic acid was transformed to DHA, its concentration being higher in MAP-stored fresh-cut spinach than in airstored samples after 1 week storage.

There is scarce information about the effects of high  $O<sub>2</sub>$  concentrations on the antioxidant content of freshcut produce. Results by different authors are somewhat contradictory. Day  $(2001)$  reported that high  $O<sub>2</sub>$  atmospheres did not result in a further decrease in ascorbic acid contents in prepared lettuce. However, it has been shown that the vitamin C content in fresh-cut pears packaged under 70 kPa  $O<sub>2</sub>$  is rapidly lost in comparison with a 2.5 kPa  $O_2$  + 7 kPa  $CO_2$  atmosphere (Oms-Oliu *et al.*, 2007b). Although the availability of  $O_2$  is the main factor affecting vitamin C degradation, high  $CO<sub>2</sub>$ levels appeared to have a negative effect on the vitamin C content of fresh-cut pears and apples (Soliva-Fortuny & Martín-Belloso, 2003b; Soliva-Fortuny et al., 2004b).

Agar *et al.* (1999) reported that high  $CO<sub>2</sub>$  levels stimulated ascorbic acid oxidation and inhibited the reduction of DHA to ascorbic acid. In fresh-cut pears stored under a 2.5 kPa  $O_2$  + 7 kPa  $CO_2$  atmosphere, a substantial loss of vitamin C after 1 week could be related to anoxic conditions reached inside the packages (Oms-Oliu et al., 2008a). According to Tudela et al.  $(2002)$ , high  $CO<sub>2</sub>$  levels in fresh-cut potatoes increased vitamin C loss by accelerating ascorbate peroxidasecatalysed oxidation processes. According to this claim, an important increase in peroxidase activity was shown during storage of fresh-cut 'Piel de Sapo' melon packaged under a 2.5 kPa  $O_2$  + 7 kPa  $CO_2$  atmosphere (Oms-Oliu et al., 2008d).

The effect of MAP on other antioxidant constituents of fruits and vegetables has been also studied. Odriozola-Serrano et al. (2008) reported a good retention of lycopene in fresh-cut tomatoes stored under 5 kPa  $O<sub>2</sub> + 5$  kPa CO<sub>2</sub> atmospheres for at least 14 days at  $5^{\circ}$ C (Fig. 2). These authors indicated that lycopene synthesis because of ripening processes, together with low oxidation of carotenoids as a result of the low availability of  $O_2$  in the package headspace, contributed to the higher levels of lycopene throughout storage. Such results were similar to those observed by Perkins-Veazie & Collins (2004) for fresh-cut watermelon stored under similar conditions for 7 days at  $5^{\circ}$ C. In addition, no losses in the total flavonoids content of fresh-cut spinach were reported throughout storage regardless of the packaging atmosphere (Gil *et al.*, 1999). These results were consistent with those previously reported on Swiss chard (Gil et al., 1998a). The stability of flavonoids contrasts with that of hydroxycinnamic acids and anthocyanins (Gil et al., 1998b). High  $O<sub>2</sub>$  atmospheres induced the loss of certain phenolic compounds in fresh-cut prepared lettuce compared with air or low  $O_2$ -modified atmospheres (Day, 2001). Cocci et al. (2006) also reported that the  $O<sub>2</sub>$  availability in the package headspace of air-stored fresh-cut apples could have led to a stronger degradation of phenolic compounds on the cut surface, which is directly exposed to molecular  $O<sub>2</sub>$  from the package headspace. A substantial loss of chlorogenic acid was reported in fresh-cut pears stored under high  $O<sub>2</sub>$  atmospheres compared with low  $O<sub>2</sub>$  environments and was associated with enzymatic browning reactions (Oms-Oliu et al., 2008a). According to these authors, the increase in other phenolic compounds such as epicatechin and quercetin in fresh-cut pears packaged under both passive modified atmospheres and high  $O<sub>2</sub>$  concentrations could be directly associated with a physiological response to stress conditions. Physiological stress may stimulate phenylalanine ammonia-lyase (PAL, EC 4.3.1.5) activity with a consequent further production of phenolic compounds, which in turn participate in subsequent tissue browning (Saltveit, 1997; Cantos et al., 2002). Research has shown that wounding increases the phenolic content of carrots (Heredia & Cisneros-Zevallos, 2002), lettuce (Kang & Saltveit, 2002) and purple-flesh potatoes (Reyes & Cisneros-Zevallos, 2003). Wound response of fresh-cut vegetables can be reduced by MAP storage. Some researchers have indicated that the use of modified atmospheres could prevent accumulation of phenolic compounds in fresh-cut produce. Beltrán et al. (2005) reported a reduction of phenolic compounds in shredded iceberg lettuce stored under reduced  $O_2$  and elevated  $CO_2$  (4 kPa  $O_2$  + 12 kPa  $CO_2$ ). They indicated a high accumulation of phenolic compounds when fresh-cut lettuce was stored in air, whereas  $CO<sub>2</sub>$  levels of >10 kPa led to a reduced phenolic content (Table 2). According to Mateos et al. (1993), exposure of fresh-cut lettuce to  $20\%$  CO<sub>2</sub> reduced total phenolic content



Figure 2 Changes in lycopene content of fresh-cut tomato from different cultivars stored under 5 kPa  $O_2$  and 5 kPa  $CO_2$ atmospheres for 21 days at 4  $°C$ . Data shown are mean  $\pm$  standard deviation. Least significant difference at  $P \leq 0.05$  (Source: Odriozola-Serrano et al., 2008).

Table 2 Changes on caffeic acid derivatives, flavonoids and total polyphenols of shredded iceberg lettuce stored in air and active modified atmosphere packaging (MAP) for 13 days at 4  $^{\circ}$ C (Source: Beltrán et al.,  $2005)^a$ 

<b>Storage</b>	Caffeic acid derivatives		<b>Flavonoids</b>		<b>Total</b> polyphenols		
days	Air	<b>MAP</b>	Air	<b>MAP</b>	Air	<b>MAP</b>	
0	17.7		3.4		21.1		
5	22.2	23.5	4.8	2.4	26.9	25.9	
9	19.0	8.7	3.3	3.1	22.3	11.8	
13	17.5	8.5	3.6	3.4	21.1	11.8	

<sup>a</sup>Mean values (three replicates) in milligram per 100 g of fresh weight. Fresh-cut lettuce was pretreated with a chlorine solution (80 mg  $\mathsf{L}^{-1}$ ). MAP: 4 kPa of  $O_2$  + 12 kPa of CO<sub>2</sub> balanced with N<sub>2</sub>.

because of a decrease in PAL activity as a result of the decrease in cytoplasmatic pH. Nevertheless, Oms-Oliu et al. (2008d) reported a higher production of phenolic compounds in fresh-cut melon stored under  $2.5$  kPa  $O<sub>2</sub>$ and 7 kPa  $CO_2$  compared with 10 kPa  $O_2$  + 7 kPa  $CO<sub>2</sub>$ , air, 20 and 70 kPa  $O<sub>2</sub>$  atmospheres, which may be related to an enhanced oxidative stress induced by toolow  $O_2$  and high  $CO_2$  concentrations (Oms-Oliu *et al.*, 2008d).

Edible coatings offer the potential to improve the nutritional quality and antioxidant properties of freshcut produce. Chien et al. (2007) maintained the ascorbic acid content of sliced red pitayas (dragon-fruit) coated with low molecular weight chitosan. Li & Barth (1998) improved carotene retention on minimally processed carrots using a cellulose-based edible coating. Oms-Oliu et al. (2008f) maintained the vitamin C and total phenolic content in pear wedges coated with alginate, gellan or pectin edible coatings containing antioxidant agents, which contributed to maintain their antioxidant potential. It was also observed that the use of pectin or alginate coatings may contribute to reduce the wounding stress induced in fresh-cut 'Piel de Sapo' melon (Oms-Oliu et al., 2008e). Serrano et al. (2006) maintained total phenolics, ascorbic acid and high retention of total antioxidant activity in table grape coated with Aloe vera gel coatings.

Edible coatings are also an excellent vehicle to enhance the nutritional value of fruits and vegetables by carrying basic nutrients that are lacking or present in low amounts in fruits and vegetables (Lin & Zhao, 2007). Despite the growing interest in incorporating nutraceutical compounds into food products, few studies have been reported suggesting the integration of bioactive ingredients into edible films or coatings to improve their functional properties. The concentration of nutrients added to the coatings must be carefully studied to know the effects on their basic functionality, namely on their barrier and mechanical properties.

Several researchers have endeavoured to incorporate minerals, vitamins and fatty acids into edible film and coating formulations to enhance the nutritional value of some fruits and vegetables. Tapia *et al.* (2008) formulated alginate and gellan-based edible coatings containing ascorbic acid for fresh-cut papaya pieces. They observed that alginate coatings appeared to perform better on ascorbic acid retention, probably because of slightly better gas barrier properties. Authors also reported that the incorporation of ascorbic acid to the coatings resulted in a substantial increase in the ascorbic acid content of the fresh-cut product, thus helping to preserve the naturally occurring amounts of this compound throughout storage. Similar results have been obtained by Ayranci & Tunc (2004) who reported that antioxidants such as citric or ascorbic acids, incorporated as an additive into a methylcellulose coating, extensively reduced ascorbic acid losses in whole apricot and peppers. Han et al. (2004) incorporated calcium and vitamin E to chitosan-based coatings to improve storability and nutritional properties of fresh and frozen strawberries and red raspberries. Their results demonstrate the capability of chitosan-based coatings to hold high concentrations of calcium or vitamin E, thus increasing significantly the content of these nutrients in both fresh and frozen fruits. For one serving (100 g), coated fruits contained about 34–59 mg of calcium and 1.7–7.7 mg of vitamin E, depending on the type of fruit and the time of storage, whereas uncoated fruits contained only 19–21 mg of calcium and 0.25–1.15 mg of vitamin E. Similarly, Hernández-Muñoz et al. (2006) observed that chitosan-coated strawberries retained more calcium (3079 g  $kg^{-1}$  dry matter) than strawberries dipped into calcium solutions (2340 g  $kg^{-1}$ ). The addition of 1% calcium gluconate to the chitosan coating formulation (1.5% in 0.5% acetic acid) did not further extend the shelf-life of strawberries. Mei et al. (2002) developed xanthan gum coatings containing high concentrations of calcium and vitamin E (5% Gluconal Cal<sup>®</sup> and 0.2%  $\alpha$ -tocopheryl acetate) with the purpose of enhancing nutritional and sensory qualities of fresh baby carrots. The results of this study showed that calcium and vitamin E contents in the coated samples increased from 2.6% to 6.6% and from 0 to about 67% of the Dietary Reference Intake values per serving (85 g) respectively, without affecting the carrots fresh aroma and flavour, sweetness, crispness and  $\beta$ -carotene levels. Because peeled baby carrots are a poor source of calcium and vitamin E, the incorporation of both into an edible coating formulation can be excellent system for enhancing the nutritional quality of this type of product. On the other hand, the addition of probiotics to obtain functional edible films and coatings has been scarcely studied. Tapia *et al.* (2007) developed the first edible films for probiotic coatings on fresh-cut apple and papaya, observing that both fruits were successfully

coated with alginate or gellan film-forming solutions containing viable bifidobacteria. In fact, values higher than  $10^6$  cfu g<sup>-1</sup> *Bifidobacterium lactis Bb*-12 were maintained for 10 days during refrigerated storage of both papaya and apple pieces. According to Samona & Robinson (1991) the count of viable bifidobacteria at the time of consumption should be of at least  $10^6$  cfu g<sup>-1</sup> to confer health benefits to the product. This work represents a promising advance in the search for new applications of edible films and coatings as carriers of diverse food additives, and opens new possibilities for the development of probiotic products.

#### Effects on flavour of fresh-cut fruits and vegetables: a possible drawback?

Modified atmosphere packaging systems can severely modify the fruit volatiles profile. Fresh-cut fruits and vegetables can develop undesirable off-odours under low  $O<sub>2</sub>$  and elevated  $CO<sub>2</sub>$  atmospheres (Soliva-Fortuny et al., 2002a, 2004a; Rojas-Graü et al., 2007b; Oms-Oliu et al., 2007a, 2007b). The beneficial effect of MAP can be attributed to a decrease of the overall metabolic activity of plant tissues. However, if  $O<sub>2</sub>$  partial pressure in MAP decreases below the fermentation threshold limit, the tissue will initiate anaerobic respiration, with the corresponding production of off-flavours and offodours (Oms-Oliu et al., 2007b). Bett et al. (2001) studied the influence of passive MAP on the flavour profile of fresh-cut 'Gala' apples. Their results indicate that overall flavour increased during the first few days after processing and then dramatically declined establishing a period of marketability of 9 days. Kim et al. (2005) found that fresh-cut lettuce packaged in films with lower  $O<sub>2</sub>$  transmission rate developed stronger offodours and more atmosphere modification than corresponding samples packages with higher  $O_2$  transmission rate. Thus, perforated and micro-perforated polymeric packages reduce off-odour formation (Izumi *et al.*, 1996). High  $O_2$  atmospheres have been also used to prevent anaerobic conditions and reduce production of fermentative metabolites (Day, 1996). However, ethanol accumulation throughout storage was detected in fresh-cut melon and pears packaged under 70 kPa  $O<sub>2</sub>$  (Oms-Oliu *et al.*, 2008b, 2008c), which was attributed to a stress response caused by the highly oxidative environment, together with the accumulation of carbon dioxide into the packages. The most accepted explanation for  $O_2$  toxicity is the formation of superoxide radicals  $(O_2^-)$ , which are destructive to cell metabolism (Fridovitch, 1975). According to Wszelaki & Mitcham (2000), the volatiles content (acetaldehyde, ethanol and ethyl acetate) of strawberry fruits stored under high  $O<sub>2</sub>$ treatments  $(≥60 \text{ kPa})$  for 14 days substantially increased compared with that observed in air-stored fruit.

Edible coatings are used as a protective barrier to reduce respiration and transpiration rates through the fruits surface. However, although reduction of gas transfer from the fruit to the environment is desirable, extremely impermeable coatings may induce anaerobic conditions that can lead to a decrease in the production of characteristic fruit aroma volatile compounds (Mattheis & Fellman, 2000; Perez-Gago et al., 2003a). Martinez-Romero et al. (2006) maintained sweet cherries coated with an Aloe vera-based coating without any detrimental effect on taste, aroma or flavours during storage.

On the other hand, it is important to highlight that many compounds used in the development of edible films and coatings, including edible matrixes, plasticisers and other active ingredients, can affect the taste and odour of coated products. These compounds have their own characteristic flavour and colour, and interaction between ingredients may generate some changes in the sensory profile. For instance, the use of chitosan-based coatings may generate slight flavour modifications because of its typical astringent⁄ bitter taste. Although, Han et al. (2005) reported that the application of a chitosan-based coating resulted in no perception of astringency in fresh strawberries. In the same way, Chien et al. (2007) observed that a chitosan coating did not influence the natural taste of sliced red pitayas. Their results contrast with those reported by Vargas et al. (2006), who observed that chitosan led to a significant decrease in the aroma and flavour of strawberries, especially when high concentrations of oleic acid were used with the purpose of increasing the moisture barrier properties.

On the other hand, the incorporation of certain antibrowning or antimicrobial agents into edible coating formulations may have detrimental consequences on the flavour of the coated product. Rojas-Graü et al. (2007a) detected a residual aromatic herbal taste in fresh-cut apples coated with an apple puree-alginate film containing a low concentration of oregano oil, which had been added with an antimicrobial purpose. Some authors have indicated that high concentrations of sulphur-containing compounds such as N-acetylcysteine and glutathione may produce an unpleasant odour in fruits and vegetables (Iyidogan & Bayindirli, 2004; Richard et al., 1992; Rojas-Graü et al., 2006). In this regard, Perez-Gago et al. (2006) detected a smell of sulphur compounds in fresh-cut apples coated with a whey protein concentrate/beeswax formulation containing cysteine as an antioxidant agent.

#### Final remarks

Traditional MAP atmospheres are rarely sufficient to ensure safety and achieve high-quality products. Most currently used MAP systems alone do not effectively prevent wounding-induced browning and deteriorative processes, which are triggered when tissue integrity is damaged as a consequence of minimal processing operations. In addition, polymeric films used in MAP have some limitations because of their structure and permeation characteristics. They may promote water loss, which results in texture changes, translucency and/or surface dehydration, or, oppositely, they can even increase the formation of water condensates that favour microbiological proliferation. Furthermore, combination of MAP systems with edible coatings can be a feasible way of improving microbial stability and quality of fresh-cut fruits and vegetables, thus extending their shelf-life. In addition, edible films and coatings offer opportunities to reduce the need for synthetic packaging materials and improve their recyclability. A new generation of edible coatings is being currently developed, allowing the incorporation and ⁄ or controlled release of active compounds using nanotechnological solutions. Nowadays, nanotechnologies are being used to enhance the nutritional aspects of food by means of nanoscale additives and nutrients and nanosised delivery systems for bioactive compounds. Incorporation of active ingredients into edible coatings for fresh-cut fruits is feasible through microencapsulation solutions or the use of novel multilayer systems (nanolaminates). These nanolaminate coatings could include various functional agents such as antimicrobials, anti-browning agents, antioxidants, enzymes, flavourings and colourants, to increase shelf-life and maintain quality of the coated foods.

#### Acknowledgments

This work was supported by the Ministry of Science and Technology (AGL2003-09208-C03-01), the Departament d'Universitats, Recerca i Societat de la Informacio´ of the Generalitat de Catalunya (Spain) and the University of Lleida (Spain).

#### References

- Agar, I.T., Massantini, R., Hess-Pierce, B. & Kader, A.A. (1999). Postharvest  $CO<sub>2</sub>$  and ethylene production and quality maintenance of fresh-cut kiwifruit slices. Journal of Food Science, 64, 433–440.
- Aguayo, E., Allende, A. & Artés, F. (2003). Keeping quality and safety of minimally fresh processed melon. European Food Research and Technology, 216, 494-499.
- Aguayo, E., Escalona, V.H. & Artés, F. (2004a). Metabolic behaviour and quality changes of whole and fresh processed melon. Journal of Food Science, 69, 148–155.
- Aguayo, E., Escalona, V. & Artés, F. (2004b). Quality of fresh-cut tomato as affected by type of cut, packaging, temperature and storage time. European Food Research and Technology, 219, 492– 499.
- Al-Ati, T. & Hotchkiss, J.H. (2002). Applications of packaging and modified atmosphere to fresh-cut fruits. In: Fresh-Cut Fruits and Vegetables: Science, Technology, and Market (edited by O. Lamikanra). Pp. 305–338. Boca Raton: CRC Press.
- Allende, A., Jacxsens, L., Devlieghere, F., Debevere, J. & Artés, F. (2002). Effect of superatmospheric oxygen packaging on sensorial quality, spoilage, and Listeria monocytogenes and Aeromonas caviae growth in fresh processed mixed salads. Journal of Food Protection, 65, 1565–1573.
- Allende, A., Luo, Y., McEvoy, J.L., Artés, F. & Wang, C.Y. (2004). Microbial and quality changes in minimally processed baby spinach leaves stored under superatmospheric oxygen and modified atmosphere conditions. Postharvest Biology and Technology, 33, 51–59.
- Amanatidou, A., Smid, E.J. & Gorris, L.G.M. (1999). Effect of elevated oxygen and carbon dioxide on the surface growth of vegetable-associated microorganisms. Journal of Applied Microbiology, 86, 429–438.
- Amanatidou, A., Slump, R.A., Gorris, L.G.M. & Smid, E.J. (2000). High oxygen and high carbon dioxide modified atmospheres for shelf-life extension of minimally processed carrots. Journal of Food Science, 65, 61–66.
- Artés, F., Conesa, M.A., Hernández, S. & Gil, M.I. (1999). Keeping quality of fresh-cut tomato. Postharvest Biology and Technology, 17, 153–162.
- Artés, F., Gómez, P.A. & Artés-Hernández, F. (2007). Physical, physiological and microbial deterioration of minimally fresh processed fruits and vegetables. Food Science and Technology International, 13, 177–188.
- Assis, O.B. & Pessoa, J.D. (2004). Preparation of thin films of chitosan for use as edible coatings to inhibit fungal growth on sliced fruits. Brazilian Journal of Food Technology, 7, 7–22.
- Avena-Bustillos, R.J., Cisneros-Zevallos, L.A., Krochta, J.M. & Saltveit, M.E. (1994a). Application of casein-lipid edible film emulsions to reduce white blush on minimally processed carrots. Postharvest Biology and Technology, 4, 319–329.
- Avena-Bustillos, R.J., Krochta, J.M., Saltveit, M.E., Rojas-Villegas, R.J. & Sauceda-Perez, J.A. (1994b). Optimization of edible coating formulations on zucchini to reduce water loss. Journal of Food Engineering, 21, 197–214.
- Avena-Bustillos, R.J., Krochta, J.M. & Saltveit, M.E. (1997). Water vapour resistance of red delicious apples and celery sticks coated with edible caseinate-acetylated monoglyceride films. Journal of Food Science, 62, 351–354.
- Ayranci, E. & Tunc, S. (2004). The effect of edible coatings on water and vitamin C loss of appricots (Armeniaca vulgaris Lam.) and green peppers (Capsicum annuum L.). Food Chemistry, 87, 339–342.
- Bai, J., Saftner, R.A., Watada, A.E. & Lee, Y.S. (2001). Modified atmosphere maintains quality of fresh-cut cantaloupe (Cucumis melo L.). Journal of Food Science, 66, 1207–1211.
- Bai, J., Saftner, R.A. & Watada, A.E. (2003). Characteristics of freshcut honeydew (*Cucumis*  $\times$  *melo* L) available to processors in winter and summer and its quality maintenance by modified atmosphere packaging. Postharvest Biology and Technology, 28, 349–359.
- Barth, M.M., Kerbel, E.L., Perry, A.K. & Schmidt, S.J. (1993). Modified atmosphere packaging affects ascorbic acid, enzyme activity and market quality of broccoli. Journal of Food Science, 58, 140–143.
- Beltrán, D., Selma, M.V., Martín, A. & Gil, M.I. (2005). Ozonated water extends the shelf life of fresh-cut lettuce. Journal of Agricultural and Food Chemistry, 53, 5654–5663.
- Bett, K.L., Ingram, D.A., Grimm, C.C. et al. (2001). Flavor of freshcut gala apples in barrier film packaging as affected by storage time. Journal of Food Quality, 24, 141-156.
- Bolin, H.R. & Huxsoll, C.C. (1991). Control of minimally processed carrot (Daucus carota) surface discoloration caused by abrasion peeling. Journal of Food Science, 56, 416–418.
- Brancoli, N. & Barbosa-Cánovas, G.V. (2000). Quality changes during refrigerated storage of packaged apple slices treated with polysaccharide films. In: Innovations in Food Processing (edited by G.V. Barbosa-Cánovas & G.W. Gould). Pp. 243-254. Pennsylvania, PA: Technomic Publishing Co.
- Cantos, E., Tudela, J.A., Gil, M.I. & Espín, J.C. (2002). Phenolic compounds and related enzymes are not rate-limiting in browning development of fresh-cut potatoes. Journal of Agricultural and Food Chemistry, 50, 3015–3023.
- Carlin, F., Nguyen-The, C., Chambroy, Y. & Reich, M. (1990). Effects of controlled atmospheres on microbial spoilage, electrolyte leakage and sugar content of fresh 'ready-to-use' carrots. International Journal of Food Science and Technology, 25, 110–119.
- Chen, C.C. & Paull, R.E. (2001). Fruit temperature and crown removal on the occurrence of pineapple fruit translucency. Scientia Horticulturae, 88, 85–95.
- Chen, M.J., Weng, Y.M. & Chen, W.L. (1999). Edible coating as preservative carriers to inhibit yeast on Taiwanese-style fruit preserves. Journal of Food Safety, 19, 89-96.
- Chien, P.J., Sheu, F. & Yang, F.H. (2007). Effects of edible chitosan coating on quality and shelf life of sliced mango fruit. Journal of Food Engineering, 78, 225–229.
- Cisneros-Zevallos, L., Salveit, M.E. & Krochta, J.M. (1995). Mechanism of surface white discoloration of peeled (minimally processed) carrots during storage. Journal of Food Science, 60, 320–323.
- Cocci, E., Rocculi, P., Romani, S. & Dalla Rosa, M. (2006). Changes in nutritional properties of minimally processed apples during storage. Postharvest Biology and Technology, 39, 265–271.
- Conway, W.S., Leverentz, W., Saftner, R., Janiciewicz, W.J., Sams, C.E. & Leblanc, E. (2000). Survival and growth of Listeria monocytogenes on fresh-cut apple slices and its interaction with Glomerella cingulata and Penicillium expansum. Plant Disease, 84, 177–181.
- Corbo, M.R., Campaniello, D., D'amato, D., Bevilacqua, A. & Sinigaglia, M. (2005). Behaviour of Listeria monocytogenes and Escherichia coli O157:H7 in fresh-sliced cactus-pear fruit. Journal of Food Safety, 25, 157–172.
- Dawson, P.L., Carl, G.D., Acton, J.C. & Han, I.Y. (2002). Effect of lauric acid and nisin-impregnated soy-based films on the growth of Listeria monocytogenes on turkey bologna. Poultry Science, 81, 721– 726.
- Day, B. (1996). High oxygen modified atmosphere packaging for fresh prepared produce. Postharvest News Information, 7, 31–34.
- Day, B. (2001). Modified atmosphere packaging of fresh fruits and vegetables – an overview. Acta Horticulturae, 553, 585–590.
- Del-Valle, V., Hernández-Muñoz, P., Guarda, A. & Galotto, M.J. (2005). Development of a cactus-mucilage edible coating  $(D$ *puntia* ficus indica) and its application to extend strawberry (Fragaria ananassa) shelf-life. Food Chemistry, 91, 751–756.
- Durango, A.M., Soares, N.F.F. & Andrade, N.J. (2006). Microbiological evaluation of an edible antimicrobial coating on minimally processed carrots. Food Control, 17, 336–341.
- Eissa, H.A.A. (2007). Effect of chitosan coating on shelf life and quality of fresh-cut mushroom. Journal of Food Quality, 30, 623–645.
- Farber, J.M. (1991). Microbiological aspects of modified-atmosphere packaging technology-a review. Journal of Food Protection, 54, 58– 70.
- Fonseca, S.C., Oliveira, F.A.R., Brecht, J.K. & Chau, K.V. (2005). Influence of low oxygen and high carbon dioxide on shredded Galega kale quality for development of modified atmosphere packages. Postharvest Biology and Technology, 35, 279–292.
- Franssen, L.R. & Krochta, J.M. (2003). Edible coatings containing natural antimicrobials for processed foods. In: Natural Antimicrobials for Minimal Processing of Foods (edited by S. Roller). Pp. 250– 262. Boca Raton: CRC Press.
- Fridovitch, I. (1975). Superoxide dismutase. Annual Reviews in Biochemistry, 44, 147–159.
- García, M.A., Martino, M.N. & Zaritzky, N.E. (1998). Starch-based coatings: effect on refrigerated strawberry (Fragaria ananassa) quality. Journal of the Science of Food and Agriculture, 76, 411–420.
- Gennadios, A. & Kurth, L.B. (1997). Application of edible coatings on meats, poultry and seafoods: a review. LWT-Food Science and Technology, 30, 337–350.
- Gil, M.I., Ferreres, F. & Tomás-Barberán, F.A. (1998a). Effect of modified atmosphere packaging on the flavonoids and vitamin C content of minimally processed Swiss chard. Journal of Agricultural and Food Chemistry, 46, 2007–2012.
- Gil, M.I., Castañer, M., Ferreres, F., Artés, F. & Tomás-Barberán, F.A. (1998b). Modified-atmosphere packaging of minimally processed ''Lollo Rosso'' (Latuca sativa). Phenolic metabolites and quality changes. European Food Research and Technology, 206, 350– 354.
- Gil, M.I., Ferreres, F. & Tomás-Barberán, F.A. (1999). Effect of postharvest storage and processing on the antioxidant constituents (flavonoids and vitamin C) of fresh-cut spinach. Journal of Agricultural and Food Chemistry, 47, 2213–2217.
- Gil, M.I., Conesa, M.A. & Artés, F. (2002). Quality changes in freshcut tomato as affected by modified atmosphere packaging. Postharvest Biology and Technology, 25, 199-207.
- Gil, M.I., Aguayo, E. & Kader, A.A. (2006). Quality changes and nutrient retention in fresh-cut versus whole fruit during storage. Journal of Agricultural and Food Chemistry, 54, 4284–4296.
- González-Aguilar, G.A., Wang, C.Y. & Buta, J.G. (2000). Maintaining quality of fresh-cut mangoes using antibrowning agents and modified atmosphere packaging. Journal of Agricultural and Food Chemistry, 48, 4202–4208.
- González-Aguilar, G.A., Ayala-Zavala, J.F., Ruiz-Cruz, S., Acedo-Félix, E. & Díaz-Cinco, M.E. (2004). Effect of temperature and modified atmosphere packaging on overall quality of fresh-cut bell peppers. LWT-Food Science and Technology, 37, 817–826.
- Gorny, J.R., Hess-Pierce, B. & Kader, A.A. (1999). Quality changes in fresh-cut peach and nectarine slices as affected by cultivar, storage atmosphere and chemical treatments. Journal of Food Science, 64, 429–432.
- Gorny, J.R., Hess-Pierce, B., Cifuente, R.A. & Kader, A.A. (2002). Quality changes in fresh-cut pear slices as affected by controlled atmospheres and chemical preservatives. Postharvest Biology and Technology, 24, 271–278.
- Gunes, G.G. & Hotchkiss, J.H. (2002). Growth and survival of Escherichia coli O157:H7 on fresh-cut apples in modified atmospheres at abusive temperatures. Journal of Food Protection, 65, 1641–1645.
- Han, C., Zhao, Y., Leonard, S.W. & Traber, M.G. (2004). Edible coatings to improve storability and enhance nutritional value of fresh and frozen strawberries (*Fragaria*  $\times$  *ananassa*) and raspberries (Rubus ideaus). Postharvest Biology and Technology, 33, 67–78.
- Han, C., Lederer, C., McDaniel, M. & Zhao, Y. (2005). Sensory evaluation of fresh strawberries (Fragaria ananassa) coated with chitosan-based edible coatings. Journal of Food Science, 70, S172– S178.
- Heimdal, H., Kuhn, B., Poll, L. & Larsen, L. (1994). Biochemical changes and sensory quality of shredded and MA-packaged iceberg lettuce. Journal of Food Science, 60, 1265–1268.
- Heredia, J.B. & Cisneros-Zevallos, L. (2002). Wounding stress on carrots increases the antioxidant capacity and the phenolics content. In: IFT Annual Meeting Book of Abstracts. Pp. 180. Chicago, IL: Institute of Food Technologists.
- Hernández-Muñoz, P., Almenar, E., Ocio, M.J. & Gavara, R. (2006). ffect of calcium dips and chitosan coatings on postharvest life of strawberries (Fragaria  $\times$  ananassa). Postharvest Biology and Technology, 39, 247–253.
- Hong, J.H. & Gross, K.C. (2000). Ethylene inhibits the development of chilling injury symptoms in fresh-cut tomato slices during cold storage. Journal of the American Society for Horticultural Science, 125, 736–741.
- Howard, L.R. & Dewi, T. (1995). Sensory, microbiological and chemical quality of mini-peeled carrots as affected by edible coating treatment. Journal of Food Science, 60, 142–144.
- Howard, L.R., Yoo, K.S., Pike, L.M. & Miller ., G.H. Jr (1994). Quality changes in diced onions stored in film packages. Journal of Food Science, 59, 110–112.
- Iyengar, R. & McEvil, A.J. (1992). Anti-browning agents alternatives to the use of sulfites in foods. Trends in Food Science and Technology, 3, 60–64.
- Iyidogan, N.F. & Bayindirli, A. (2004). Effect of L-cysteine, kojic acid and 4-hexylresorcinol combination on inhibition of enzymatic browning in Amasya apple juice. Journal of Food Engineering, 62, 299–304.
- Izumi, H., Watada, A.E. & Douglas, W. (1996). Optimum  $O_2$  or  $CO_2$ atmosphere for storing broccoli florets at various temperatures. Journal of the American Society of Horticultural Science, 121, 127-131.
- Jacxsens, L., Devlieghere, F., Van der Steen, C. & Debevere, J. (2001). Effect of high oxygen atmosphere packaging on microbial growth and sensorial qualities of fresh-cut produce. International Journal of Food Microbiology, 71, 197–210.
- Jacxsens, L., Devlieghere, F., Van der Steen, C., Siro, I. & Debevere, J. (2003). Application of ethylene absorbers in combination with high oxygen atmospheres for the storage of strawberries and raspberries. Acta Horticulturae, 600, 311–318.
- Jasso de Rodríguez, D., Hernández-Castillo, D., Rodríguez-García, R. & Angulo-Sánchez, J.L. (2005). Antifungal activity in vitro of aloe vera pulp and liquid fraction against plant pathogenic fungi. Industrial Crops and Products, 21, 81–87.
- Kader, A.A. & Ben-Yehoshua, S. (2000). Effects of superatmospheric oxygen levels on postharvest physiology and quality of fresh fruits and vegetables. Postharvest Biology and Technology, 20, 1–13.
- Kang, J.S. & Lee, D.S. (1997). Susceptibility of minimally processed green pepper and cucumber to chilling injury by apparent respiration rate. International Journal of Food Science and Technology, 32, 421–426.
- Kang, H. & Saltveit, M.E. (2002). Antioxidant capacity of lettuce leaf tissue increases after wounding. Journal of Agricultural and Food Chemistry, 50, 7536–7541.
- Ke, D. & Saltveit, M. (1989). Wound-induced ethylene production, phenolic metabolism and susceptibility to russet spotting in iceberg lettuce. Physiology Plantarum, 76, 412–418.
- Kim, J.G., Luo, Y., Tao, Y., Saftner, R.A. & Gross, K.C. (2005). Effect of initial oxygen concentration and film oxygen transmission rate on the quality of fresh-cut romaine lettuce. Journal of the Science and Agriculture, 85, 1622-1630.
- Lamikanra, O., Chen, J.C., Banks, D. & Hunter, P.A. (2000). Biochemical and microbial changes during the storage of minimally processed cantaloupe. Journal of Agricultural and Food Chemistry, 48, 5955–5961.
- Lana, M.M., Hogenkamp, M. & Koehorst, R.B.M. (2006). Application of Kubelka–Munk analysis to the study of translucency in fresh-cut tomato. Innovative Food Science and Emerging Technologies, 7, 302–308.
- Larson, A.E. & Johnson, E.A. (1999). Evaluation of botulinal toxin production in packaged fresh-cut cantaloupe and honeydew melon. Journal of Food Protection, 62, 948-952.
- Le Tien, C., Vachon, C., Mateescu, M.A. & Lacroix, M. (2001). Milk protein coatings prevent oxidative browning of apples and potatoes. Journal of Food Science, 66, 512-516.
- Lee, J.Y., Park, H.J., Lee, C.Y. & Choi, W.Y. (2003). Extending shelflife of minimally processed apples with edible coatings and antibrowning agents. LWT-Food Science and Technology, 36, 323–329.
- Li, P. & Barth, M.M. (1998). Impact of edible coatings on nutritional and physiological changes in lightly processed carrots. Postharvest Biology and Technology, 14, 51–60.
- Limbo, S. & Piergiovanni, L. (2006). Shelf life of minimally processed potatoes Part 1. Effect of high oxygen partial pressures in combination with ascorbic and citric acids on enzymatic browning. Postharvest Biology and Technology, 39, 254–264.
- Lin, D. & Zhao, Y. (2007). Innovations in the development and application of edible coatings for fresh and minimally processed fruits and vegetables. Comprehensive Reviews in Food Science and Food Safety, 6, 60–75.
- Lipton, W. (1987). Carbon dioxide-induced injury of romaine lettuce stored in controlled atmospheres. HortScience, 22, 461-463.
- López-Gálvez, G., Saltveit, M. & Cantwell, M. (1996). The visual quality of minimally processed lettuces stored in air or controlled atmosphere with emphasis on romaine and iceberg types. Postharvest Biology and Technology, 8, 179–190.
- Maftoonazad, N. & Ramaswamy, H.S. (2004). Postharvest shelf-life extension of avocados using methyl cellulose-based coating. LWT-Food Science and Technology, 38, 617–624.
- Martín-Belloso, O., Soliva-Fortuny, R. & Oms-Oliu, G. (2007). Freshcut fruits. In: Handbook of Food Products Manufacturing. Principles, Bakery, Beverages, Cereals, Cheese, Confectionary, Fats, Fruits, and Functional Foods (edited by Y.H. Hui). Pp. 879–899. New Jersey, NJ: John Wiley & Sons, Inc.
- Martinez, M.V. & Whitaker, J.R. (1995). The biochemistry and control of enzymatic browning. Trends in Food Science and Technology, 6, 195–200.
- Martínez-Romero, D., Serrano, M., Valero, D. & Castillo, S. (2003). Aplicación de Aloe vera como recubrimiento sobre frutas y hortalizas. Spain patent: 200302937.
- Martínez-Romero, D., Alburquerque, N., Valverde, J.M. et al. (2006). Postharvest sweet cherry quality and safety maintenance by Aloe Vera treatment: a new edible coating. Postharvest Biology and Technology, 39, 93–100.
- Mateos, M., Ke, D., Cantwell, M. & Kader, A.A. (1993). Phenolic metabolism and ethanolic fermentation of intact and cut lettuce exposed to CO<sub>2</sub>-enriched atmospheres. Postharvest Biology and Technology, 3, 225–233.
- Mattheis, J. & Fellman, J.K. (2000). Impacts of modified atmosphere packaging and controlled atmospheres on aroma, flavor, and quality of horticultural commodities. HortTechnology, 10, 507–510.
- McHugh, T.H. & Senesi, E. (2000). Apple wraps: a novel method to improve the quality and extend the shelf life of fresh-cut apples. Journal of Food Science, 65, 480–485.
- Mei, Y., Zhao, Y., Yang, J. & Furr, H.C. (2002). Using edible coating to enhance nutritional and sensory qualities of baby carrots. Journal of Food Science, 67, 1964–1968.
- Min, S. & Krochta, J.M. (2005). Inhibition of Penicillium commune by edible whey protein films incorporating lactoferrin, lactoferrin hydrolysate, and lactoperoxidase systems. Journal of Food Science, 70, M87–M94.
- Montero-Calderón, M., Rojas-Graü, M.A. & Martin-Belloso, O. (2008). Effect of packaging conditions on quality and shelf-life of fresh-cut pineapple (Ananas comosus). Postharvest Biology and Technology, 50, 182–189.
- Nisperos-Carriedo, M.O., Baldwin, E.A. & Shaw, P.E. (1992). Development of an edible coating for extending postharvest life of selected fruits and vegetables. Florida State Horticulture Society, 104, 122–125.
- O'Connor-Shaw, R., Roberts, R., Ford, A. & Nottingham, S. (1994). Shelf-life of minimally processed honeydew, kiwifruit, papaya, pineapple and cantaloupe. Journal of Food Science, 59, 1202–1206.
- Odriozola-Serrano, I., Soliva-Fortuny, R. & Martín-Belloso, O. (2008). Effect of minimal processing on bioactive compounds and colour attributes of fresh-cut tomatoes. LWT-Food Science and Technology, 41, 217–226.
- Olivas, G.I., Rodriguez, J.J. & Barbosa-Cánovas, G.V. (2003). Edible coatings composed of methylcellulose, stearic acid, and additives to preserve quality of pear wedges. Journal of Food Processing and Preservation, 27, 299-320.
- Olivas, G.I., Mattinson, D.S. & Barbosa-Canovas, G.V. (2007). Alginate coatings for preservation of minimally processed 'Gala' apples. Postharvest Biology and Technology, 45, 89-96.
- Oms-Oliu, G., Soliva-Fortuny, R. & Martín-Belloso, O. (2007a). Effect of ripeness on the shelf-life of fresh-cut melon preserved by modified atmosphere packaging. European Food Research and Technology, 225, 301–311.

#### 888 The use of packaging techniques to maintain freshness in fresh-cut fruits and vegetables  $M^a$ . A. Rojas-Graü et al.

- Oms-Oliu, G., Soliva-Fortuny, R. & Martín-Belloso, O. (2007b). Respiratory rate and quality changes in fresh-cut pears as affected by superatmospheric oxygen. Journal of Food Science, 72, 456– 463.
- Oms-Oliu, G., Odriozola-Serrano, I., Soliva-Fortuny, R. & Martín-Belloso, O. (2008a). Antioxidant content of fresh-cut pears stored in high-O2 active packages compared with conventional low-O2 active and passive modified atmosphere packaging. Journal of Agricultural and Food Chemistry,  $56$ ,  $932-940$ .
- Oms-Oliu, G., Soliva-Fortuny, R. & Martín-Belloso, O. (2008b). Physiological and microbiological changes in fresh-cut pears stored in high oxygen active packages compared with low oxygen active and passive modified atmosphere packaging. Postharvest Biology and Technology,  $48$ ,  $295-301$ .
- Oms-Oliu, G., Raybaudi-Massilia Martínez, R.M., Soliva-Fortuny, R. & Martín-Belloso, O. (2008c). Effect of superatmospheric and low oxygen modified atmospheres on shelf-life extension of fresh-cut melon. Food Control, 19, 191–199.
- Oms-Oliu, G., Odriozola-Serrano, I. & Soliva-Fortuny, R. (2008d). The role of peroxidase on the antioxidant potential of fresh-cut 'Piel de Sapo' melon packaged under different modified atmospheres. Food Chemistry, 106, 1085–1092.
- Oms-Oliu, G., Soliva-Fortuny, R. & Martín-Belloso, O. (2008e). Using polysaccharide-based edible coatings to enhance quality and antioxidant properties of fresh-cut melon. LWT-Food Science and Technology, 41, 1862–1870.
- Oms-Oliu, G., Soliva-Fortuny, R. & Martín-Belloso, O. (2008f). Edible coatings with antibrowning agents to maintain sensory quality and antioxidant properties of fresh-cut pears. Postharvest Biology and Technology, 50, 87–94.
- Palmer-Wright, K. & Kader, A.A. (1997). Effect of controlledatmosphere storage on the quality and carotenoid content of sliced persimmons and peaches. Postharvest Biology and Technology, 10,  $89 - 97$
- Park, S.I., Stan, S.D., Daeschel, M.A. & Zhao, Y.Y. (2005). Antifungal coatings on fresh strawberries (*Fragaria*  $\times$  *ananassa*) to control mold growth during cold storage. Journal of Food Science, 70, M202–M207.
- Pen, L.T. & Jiang, Y.M. (2003). Effects of chitosan coating on shelf life and quality of fresh-cut Chinese water chestnut. LWT-Food Science and Technology, 36, 359–364.
- Perez-Gago, C., Rojas, C. & Del Rio, M.A. (2003a). Effect of hydroxypropyl methylcellulose-lipid edible composite coating on plum (cv. Autumn giant) quality during storage. Journal of Food Science, 68, 879–883.
- Perez-Gago, M.B., Serra, M., Alonso, M., Mateos, M. & Del Rio, M.A. (2003b). Effect of solid content and lipid content of whey protein isolate-beeswax edible coatings on color change of fresh-cut apples. Journal of Food Science, 68, 2186–2191.
- Perez-Gago, M.B., Serra, M., Alonso, M., Mateos, M. & Del Rio, M.A. (2005). Effect of whey protein- and hydroxypropyl methylcellulose-based edible composite coatings on color change of fresh-cut apples. Postharvest Biology and Technology, 36, 77–85.
- Perez-Gago, M.B., Serra, M. & Del Rio, M.A. (2006). Color change of fresh-cut apples coated with whey protein concentrate-based edible coatings. Postharvest Biology and Technology, 39, 84–92.
- Perkins Veazie, P.M. & Collins, J.K. (2004). Carotenoid changes in stored watermelon fruit. Hort Science, 39, 667.
- Petersen, K., Nielsen, P.V., Bertelsen, G. et al. (1999). Potential of biobased materials for food packaging. Trends in Food Science and Technology, 10, 52–68.
- Portella, S. & Cantwell, M. (1998). Quality changes of minimally processed honeydew melons stored in air or controlled atmosphere. Postharvest Biology and Technology, 14, 351–357.
- Poubol, J. & Izumi, H. (2005). Physiology and microbiological quality of fresh-cut mango cubes as affected by high-O2 controlled atmospheres. Journal of Food Science, 70, 286–291.
- Qi, L., Wu, T. & Watada, A.E. (1999). Quality changes of fresh-cut honeydew melons during controlled atmosphere storage. Journal of Food Quality, 22, 513–521.
- Rattanapanone, N., Lee, Y. & Watada, A.E. (2001). Quality and microbial changes of fresh-cut mango cubes held in controlled atmosphere. HortScience, 36, 1091–1095.
- Raybaudi-Massilia, R.M., Rojas-Graü, M.A., Mosqueda-Melgar, J. & Martín-Belloso, O. (2008a). Comparative study on essential oils incorporated into an alginate-based edible coating to assure the safety and quality of fresh-cut Fuji apples. Journal of Food Protection, 71, 1150–1161.
- Raybaudi-Massilia, R.M., Mosqueda-Melgar, J. & Martín-Belloso, O. (2008b). Edible alginate-based coating as carrier of antimicrobials to improve shelf-life and safety of fresh-cut melon. International Journal of Food Microbiology, 121, 313–327.
- Reyes, L.F. & Cisnero-Zevallos, L. (2003). Wounding stress increases the phenolic content and antioxidant capacity of purple-flesh potatoes. Journal of Agricultural and Food Chemistry, 51, 5296–5300.
- Richard, F.C., Goupy, P.M. & Nicolas, J.J. (1992). Cysteine as an inhibitor of enzymatic browning. 2. Kinetic studies. Journal of Agricultural and Food Chemistry, 40, 2108–2114.
- Rojas-Graü, M.A. & Martín-Belloso, O. (2008b). Current advances in quality maintenance of fresh-cut fruits. Stewart Postharvest Review, 2, 6.
- Rojas-Graü, M.A., Sobrino-López, A., Tapia, M.S. & Martín-Belloso, O. (2006). Browning inhibition in fresh-cut 'Fuji' apple slices by natural antibrowning agents. Journal of Food Science, 71, S59–S65.
- Rojas-Graü, M.A., Raybaudi-Massilia, R.M., Soliva-Fortuny, R.C., Avena-Bustillos, R.J., McHugh, T.H. & Martín-Belloso, O. (2007a). Apple puree-alginate edible coating as carrier of antimicrobial agents to prolong shelf-life of fresh-cut apples. Postharvest Biology and Technology, 45, 254–264.
- Rojas-Graü, M.A., Grasa-Guillem, R. & Martín-Belloso, O. (2007b). Quality changes in fresh-cut Fuji apple as affected by ripeness stage, antibrowning agents, and stored atmosphere. Journal of Food Science, 72, 36–43.
- Rojas-Graü, M.A., Tapia, M.S., Rodríguez, F.J., Carmona, A.J. & Martín-Belloso, O. (2007c). Alginate and gellan based edible coatings as support of antibrowning agents applied on fresh-cut Fuji apple. Food Hydrocolloids, 21, 118–127.
- Rojas-Graü, M.A., Soliva-Fortuny, R. & Martín-Belloso, O. (2008a). Effect of natural antibrowning agents on color and related enzymes in fresh-cut Fuji apples as an alternative to the use of ascorbic acid. Journal of Food Science, 73, 267–272.
- Romanazzi, G., Nigro, F., Ippolito, A., Di Venere, D. & Salerno, M. (2002). Effects of pre- and postharvest chitosan treatments to control storage grey mold of table grapes. Journal of Food Science, 67, 1862–1867.
- Saftner, R.A., Abbott, J.A., Bhagwat, A.A. & Vinyard, B.T. (2005). Quality measurement of intact and fresh-cut slices of Fuji, Granny Smith, Pink Lady, and GoldRush apples. Journal of Food Science, 70, S317–S324.
- Saladié, M., Matas, A.J., Isaacson, T. et al. (2007). A re-evaluation of the key factors that influence tomato fruit softening and integrity. Plant Physiology, 144, 1012–1028.
- Saltveit, M.E. (1997). Physical and physiological changes in minimally processed fruits and vegetables. In: Phytochemistry of Fruit and Vegetables. Proceedings of the Phytochemical Society of Europe (edited by F.A. Tomás-Barberán & R.J. Robins). Pp. 205–220. New York, NY: Oxford University.
- Samona, A. & Robinson, R.K. (1991). Enumeration of Bifidobacteria in dairy products. Journal of the Society of Dairy Technology, 44, 64–66.
- Sebti, I., Ham-Pichavant, F. & Coma, V. (2002). Edible bioactive fatty acid-cellulosic derivative composites used in food-packaging applications. Journal of Agricultural and Food Chemistry, 50, 4290–4294.
- Serrano, M., Valverde, J.M., Guillén, F., Castillo, S., Martínez-Romero, D. & Valero, D. (2006). Use of aloe vera gel coating preserves the functional properties of table grapes. Journal Agricultural and Food Chemistry, 54, 3882-3886.
- Shon, J. & Haque, Z.U. (2007). Efficacy of sour whey as a shelf-life enhancer: use in antioxidative edible coatings of cut vegetables and fruit. Journal of Food Quality, 30, 581–593.
- Soliva-Fortuny, R.C. & Martín-Belloso, O. (2003a). New advances in extending the shelf-life of fresh-cut fruits: a review. Trends in Food Science and Technology, 14, 341–353.
- Soliva-Fortuny, R.C. & Martín-Belloso, O. (2003b). Microbiological and biochemical changes in minimally processed fresh-cut conference pears. European Food Research and Technology, 217, 4–9.
- Soliva-Fortuny, R.C., Oms-Oliu, G. & Martín-Belloso, O. (2002a). Effects of ripeness stages on the storage atmosphere, colour, and textural properties of minimally processed apple slices. Journal of Food Science, 67, 1958–1963.
- Soliva-Fortuny, R.C., Grigelmo-Miguel, N., Hernando, I., Lluch, M.A. & Martín-Belloso, O. (2002b). Effect of minimal processing on the textural and structural properties of fresh-cut pears. Journal of the Science of Food and Agriculture, 82, 1682–1688.
- Soliva-Fortuny, R.C., Lluch, M.A., Quiles, A., Grigelmo-Miguel, N. & Martín-Belloso, O. (2002c). Evaluation of textural properties and microstructure during storage of minimally processed apples. Journal of Food Science, 68, 312-317.
- Soliva-Fortuny, R.C., Alós-Saiz, N., Espachs-Barroso, A. & Martín-Belloso, O. (2004a). Influence of maturity at processing on quality attributes of fresh-cut 'Conference' pears. Journal of Food Science, 69, 290–294.
- Soliva-Fortuny, R.C., Elez-Martínez, P. & Martín-Belloso, O. (2004b). Microbiological and biochemical stability of fresh-cut apples preserved by modified atmosphere packaging. Innovative Food Science and Emerging Technologies, 5, 215–224.
- Sonti, S., Prinyawiwatkul, W., No, H.K. & Janes, M.E. (2003). Maintaining quality of fresh-cut apples with edible coating during 13-days refrigerated storage. In: IFT Annual Meeting Book of Abstracts. Pp. 45. New Orleans, LA: Institute of Food Technologists.
- Sothornvit, R. & Rodsamran, P. (2006). Effect of a mango film on quality of whole and minimally processed mangoes. Postharvest Biology and Technology, 47, 407–415.
- Szabo, E.A., Scurrah, K.J. & Burrows, J.M. (2000). Survey for psychrotrophic bacterial pathogens in minimally processed lettuce. Letters of Applied Microbiology, 30, 456–460.
- Tapia, M.S., Rodríguez, F.J., Rojas-Graü, M.A. & Martín-Belloso, O. (2005). Formulation of alginate and gellan based edible coatings with antioxidants for fresh-cut apple and papaya. In: IFT Annual

Meeting Book of Abstracts. Pp. 36. New Orleans, LA: Institute of Food Technologists.

- Tapia, M.S., Rojas-Graü, M.A., Rodríguez, F.J., Ramírez, J., Carmona, A. & Martín-Belloso, O. (2007). Alginate- and Gellanbased edible films for probiotic coatings on fresh-cut fruits. Journal of Food Science, 72, E190–E196.
- Tapia, M.S., Rojas-Graü, M.A., Carmona, A., Rodriguez, F.J., Soliva-Fortuny, R. & Martín-Belloso, O. (2008). Use of alginate and gellan-based coatings for improving barrier, texture and nutritional properties of fresh-cut papaya. Food Hydrocolloids, 22, 1493–1503.
- Toivonen, P.M.A. & Brummell, D.A. (2008). Biochemical bases of appearance and texture changes in fresh-cut fruit and vegetables. Postharvest Biology and Technology, 48, 1–14.
- Tudela, J.A., Espín, J.C. & Gil, M.I. (2002). Vitamin C retention in fresh-cut potatoes. Postharvest Biology and Technology, 26, 75–84.
- Valverde, J.M., Valero, D., Martinez-Romero, D., Guillen, F., Castillo, S. & Serrano, M. (2005). Novel edible coating based on Aloe vera gel to maintain table grape quality and safety. Journal of Agricultural and Food Chemistry, 53, 7807–7813.
- Van der Steen, C., Jacxsens, L., Devlieghere, F. & Debevere, J. (2002). Combining high oxygen atmospheres with low oxygen modified atmosphere packaging to improve the keeping quality of strawberries and raspberries. Postharvest Biology and Technology, 26, 49–58.
- Vargas, M., Albors, A., Chiralt, A. & Gonzalez-Martinez, C. (2006). Quality of cold-stored strawberries as affected by chitosan-oleic acid edible coatings. Postharvest Biology and Technology, 41, 164– 171.
- Vilas-Boas, E. & Kader, A.A. (2006). Effect of atmospheric modfiication, 1-MCP and chemicals on quality of fresh-cut banana. Postharvest Biology and Technology, 39, 155–162.
- Watada, A.E., Ko, N.P. & Minott, D.A. (1996). Factors affecting quality of fresh-cut horticultural products. Postharvest Biology and Technology, 9, 115–125.
- Wong, W.S., Tillin, S.J., Hudson, J.S. & Pavlath, A.E. (1994). Gas exchange in cut apples with bilayer coatings. Journal of Agricultural and Food Chemistry, 42, 2278–2285.
- Wszelaki, A.L. & Mitcham, E.J. (2000). Effects of supeatmospheric oxygen on strawberry fruit quality and decay. Postharvest Biology and Technology, 20, 125–133.
- Yang, S.F. & Hoffman, N.E. (1984). Ethylene biosynthesis and its regulation in higher plants. Annual Review of Plant Physiology, 35, 155–189.
- Zhuang, R., Beuchat, L.R., Chinnan, M.S., Shewfelt, R.L. & Huang, Y.W. (1996). Inactivation of Salmonella montevideo on tomatoes by applying cellulose-based films. Journal of Food Protection, 59, 808– 812.